

Evaluating heterogeneity in ASD symptomatology, cognitive ability, and adaptive functioning among 16p11.2 CNV carriers

Caitlin M. Hudac , Joanna Bove, Shelley Barber, Michael Duyzend, Ari Wallace, Christa Lese Martin, David H. Ledbetter, Ellen Hanson, Robin P. Goin-Kochel , LeeAnne Green-Snyder, Wendy K. Chung, Evan E. Eichler, and Raphael A. Bernier

Individuals with 16p11.2 copy number variant (CNV) show considerable phenotypic heterogeneity. Although autism spectrum disorder (ASD) is reported in approximately 20–23% of individuals with 16p11.2 CNVs, ASD-associated symptoms are observed in those without a clinical ASD diagnosis. Previous work has shown that genetic variation and prenatal and perinatal birth complications influence ASD risk and symptom severity. This study examined the impact of genetic and environmental risk factors on phenotypic heterogeneity among 16p11.2 CNV carriers. Participants included individuals with a 16p11.2 deletion ($N = 96$) or duplication ($N = 77$) with exome sequencing from the Simons VIP study. The presence of prenatal factors, perinatal events, additional genetic events, and gender was studied. Regression analyses examined the contribution of each risk factor on ASD symptomatology, cognitive functioning, and adaptive abilities. For deletion carriers, perinatal and additional genetic events were associated with increased ASD symptomatology and decrements in cognitive and adaptive functioning. For duplication carriers, secondary genetic events were associated with greater cognitive impairments. Being female sex was a protective factor for both deletion and duplication carriers. Our findings suggest that ASD-associated risk factors contribute to the variability in symptom presentation in individuals with 16p11.2 CNVs. *Autism Res* 2020, 13: 1300–1310. © 2020 International Society for Autism Research, Wiley Periodicals, LLC.

Lay Summary: There are a wide range of autism spectrum disorder (ASD) symptoms and abilities observed for individuals with genetic changes of the 16p11.2 region. Here, we found perinatal complications contributed to more severe ASD symptoms (deletion carriers) and additional genetic mutations contributed to decreased cognitive abilities (deletion and duplication carriers). A potential protective factor was also observed for females with 16p11.2 variations.

Keywords: 16p11.2 deletion; 16p11.2 duplication; autism spectrum disorder; individual variability/heterogeneity; cognitive functioning; adaptive functioning

Introduction

Copy number variations (CNVs) in the ~600 kb BP4-BP5 region of 16p11.2 are associated with autism spectrum disorder (ASD), schizophrenia, mood disorders, attention deficit and hyperactivity disorder, intellectual disability, language disorders, motor disorders, obesity, and other neurodevelopmental disorders [Niarchou et al., 2019a; Niarchou et al., 2019b; Kumar et al., 2008; McCarthy et al., 2009; Weiss et al., 2008; Zufferey et al., 2012]. For subjects with a psychiatric or language disorder, frequency estimates are 0.1% and 0.04% for 16p11.2

deletions and duplications, respectively, while accounting for approximately 1% of those clinically diagnosed with ASD. 16p11.2 CNVs are also present and potentially asymptomatic in about 0.01–0.043% of the general population, respectively [Rosenfeld, Coe, Eichler, Cuckle, & Shaffer, 2013; Kirov et al., 2014]. With 16p11.2 CNV carriers, ASD rates are estimated at approximately 25% for deletion carriers and 20% for duplication carriers [Hanson et al., 2015; Snyder et al., 2016]. Within deletion carriers, quantitative assessment across phenotypic domains indicates clinician-observed autism symptom severity ranges from within normal range to severely impaired, with

From the Department of Psychiatry and Behavioral Sciences, University of Washington, Seattle, Washington, USA (C.M.H., R.A.B.); Department of Medicine, University of Washington, Seattle, Washington, USA (J.B.); Department of School Psychology, University of Washington, Seattle, Washington, USA (S.B., A.W.); Department of Genome Sciences, University of Washington, Seattle, Washington, USA (M.D., E.E.E.); Autism and Developmental Medicine Institute, Geisinger, Danville, Pennsylvania, USA (C.L.M., D.H.L.); Developmental Medicine, Children's Hospital Boston/Harvard Medical School, Boston, Massachusetts, USA (E.H.); Department of Pediatrics, Baylor College of Medicine, Houston, Texas, USA (R.P.G.-K.); Simons Foundation, New York, New York, USA (L.G.-S.); Department of Pediatrics, Columbia University Irving Medical Center, New York, New York, USA (W.K.C.); Department of Medicine, Columbia University Irving Medical Center, New York, New York, USA (W.K.C.); Center for Youth Development and Intervention and Department of Psychology at University of Alabama, Tuscaloosa, Alabama, USA (C.M.H.)

Caitlin M. Hudac and Joanna Bove contributed equally as first authors.

Received December 16, 2019; accepted for publication June 3, 2020

Address for correspondence and reprints: Caitlin M. Hudac, Center for Youth Development and Intervention, Department of Psychology, University of Alabama, Box 870348, Tuscaloosa, AL 35401. E-mail: cmhudac@ua.edu

Published online 28 June 2020 in Wiley Online Library (wileyonlinelibrary.com)

DOI: 10.1002/aur.2332

© 2020 International Society for Autism Research, Wiley Periodicals, LLC.

comparable variability indicated from parent report [Hanson et al., 2015; Moreno-De-Luca et al., 2015]. A similar pattern of significant ASD-symptom severity variability is observed in duplication carriers [Snyder et al., 2016].

ASD is a behaviorally defined neurodevelopmental disorder characterized by deficits in social communication and interaction and the presence of restrictive and repetitive behaviors (RRBs). Converging evidence provides strong support for the genetic contribution to ASD with hundreds of genomic loci, including 16p11.2, and genes implicated [Coe et al., 2019; Iossifov et al., 2014; Sanders et al., 2015]. Furthermore, the presence of “secondary hits” and multiple genetic risk factors is associated with increased impairment for individuals with ASD [Guo et al., 2018; Girirajan et al., 2012; O’Roak et al., 2011; Samocha et al., 2014]. Studies examining environmental risk factors of ASD have revealed significant associations between prenatal and perinatal risk factors and ASD symptomatology, which may confer independent risk of ASD symptomatology, be the sequelae of genetic insults, or function interactively with other risk factors [Hisle-Gorman et al., 2018; Chien et al., 2019; Gardener, Spiegelman, & Buka, 2011; Langridge et al., 2013; Matelski & Van de Water, 2016; Visser et al., 2013]. Prenatal events such as advanced parental age, preeclampsia, infection or autoimmune disease during pregnancy, and the use of psychotropic or antidepressant medications have been associated with an increased risk of ASD and other neurodevelopmental disorders [Chen et al., 2016; Gardener, Spiegelman, & Buka, 2009; Jiang et al., 2016; Kaplan, Keskin-Arslan, Acar, & Sozmen, 2016; Rais & Rais, 2014; Dachew, Mamun, Maravilla, & Alati, 2018]. Perinatal events such as abnormal (breech) presentation, low birthweight, preterm birth, low 5-min Apgar score, and respiratory distress have also been associated with an increased risk of ASD [Gardener et al., 2011; Modabbernia, Mollon, Boffetta, & Reichenberg, 2016]. Investigations examining interactions between genetic risk and a prenatal risk factor, maternal infection during pregnancy, and increased ASD severity suggest an additive effect of multiple events on the severity of ASD symptomatology [Mazina et al., 2014], across a wide variety of CNV loci.

The study of variability in the phenotypic expression of 16p11.2 implicates the presence of secondary CNVs [Duyzend et al., 2016] and contributions of family background to the observed variability [D’Angelo et al., 2016; Moreno-De-Luca et al., 2015]. In ASD more broadly, pre- and peri-natal factors have been proposed as risk factors in the development of ASD symptoms [Gardener et al., 2009; Gardener et al., 2011; Langridge et al., 2013] and examined as predictors for variability in autism symptomatology [Matelski & Van de Water, 2016; Perrone-McGovern, Simon-Dack, & Niccolai, 2015; Schrieken et al., 2013]. Similarly, within ASD the phenotype varies with sex; females with an ASD diagnosis show less severe

symptomatology relative to males with ASD and are less likely to be diagnosed with ASD than males, suggesting a female-protective effect [Krumm et al., 2015; Park et al., 2016; Polyak, Rosenfeld, & Girirajan, 2015]. These reported gender differences in the phenotypic presentation have been associated with secondary genetic hits [Jacquemont et al., 2014] and differences in recognition and diagnostic practices [Constantino & Charman, 2012; Kreiser & White, 2014; Lai, Lombardo, Auyeung, Chakrabarti, & Baron-Cohen, 2015].

To expand on these studies of 16p11.2 and incorporate findings from the literature within the field of ASD, we investigated the impact of other genetic and environmental factors that could predict phenotypic heterogeneity among 16p11.2 deletion and duplication carriers. Specifically, we examined the contribution of secondary genetic events, pre- and peri-natal events, and sex on the variability of ASD symptomatology. We focused on ASD symptomatology given the consistent findings of high rates of ASD diagnosis within this population, as well as consistent social-communication deficits and reports of repetitive interests and behaviors in individuals not meeting diagnostic criteria [Hanson et al., 2015; Snyder et al., 2016].

Methods

Participants

Prior to additional genetic screening, subjects included individuals with the recurrent 600 kb BP4–BP5 16p11.2 duplication or deletion participating in the Simons Variation in Individuals Project [Simons-VIP; Simons VIP Consortium, 2012]. All participants were fluent in English and did not have other genetic diagnoses, additional clinically recognized CNVs, or severe neurological insults. Identified probands and their family members received cascade genetic testing, yielding an initial population size of 137 deletion carriers and 129 duplication carriers (final population characterized is shown in Table 2). See Simons VIP Consortium [2012] for data collection methods. As part of participation in the consortium, informed consent was obtained for all participants in accordance with the ethical standards of each local institutional research review board and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

Measures

Additional genetic factors. Sequencing to identify additional rare CNVs was conducted using Illumina HumanOmniExpress microarray platforms [see Duyzend et al., 2016 for methods]. Exome sequencing data from the Simons Variation in Individuals Project were utilized to assess genome-wide exonic variation (as described on

SFARI website; <http://sfari.org>). We analyzed a total of 431 exomes from the Simons VIP. We applied several filters to the variants present in the exome VCF available through SFARI base using the VCFLib [Garrison, 2012 VCFLib] and VCFtools [Danecek et al., 2011] software suites. For filtering we selected only variants with a PASS flag, depth >10, QUAL >20, removed dbSNP sites excluding sites after build 129, removed Mills and 1,000 genomes gold standard indels, removed segmental duplications and tandem repeats, removed variants found at >1% frequency in dbSNP144, and removed sex chromosomes. We annotated using SeattleSeq build 138, selected the transcript with the most severe annotation for the variant, and retained only non-synonymous and splice variants. We additionally annotated each variant with the Combined Annotation Dependent Depletion (CADD) v1.3 score [Kircher et al., 2014], the allele frequency from the Exac database containing nonpsychiatric cases (ftp://ftp.broadinstitute.org/pub/ExAC_release/release0.3/subsets/ExAC.r0.3.nonpsych.sites.vcf.gz), and the Residual Variation Intolerance Score [Petrovski, Wang, Heinzen, Allen, & Goldstein, 2013; version 3_12_Mar16, columns all 0.1% and %All 0.1%] Tables S1–S3 summarize CNV and SNV results for individuals with sequencing data.

Sequencing results yielded a final study population of 96 deletion carriers and 77 duplication carriers. The presence of additional rare CNVs [defined as occurring with <0.1% frequency in control individuals in the Duyzend et al., 2016 study] and deleterious SNVs (defined as an SNV with a CADD score of >20) were tallied for each individual. Additional SNVs with unknown significance (CADD score = <20) are included in Table S3, but these variants were not included in the frequency of SNVs. Each individual was categorized as having 1 or more additional CNVs, SNVs, both, or none.

Prenatal and perinatal complications. Prenatal risk factors and perinatal events were identified for each participant through parent/caregiver interview. These events were identified as having an increased risk for ASD diagnosis based on findings from meta-analyses conducted by Gardener et al. [2009] and Gardener et al. [2011] for prenatal and perinatal events that demonstrated a positive association with ASD diagnosis or symptomatology. Table 1 lists the prenatal risk factors and perinatal events that were collected in the Simons VIP sample and selected for analysis. The number of events was calculated for each individual yielding a single value ranging from zero to nine.

Phenotypic measurement. ASD symptomatology was based on scores obtained from the Autism Diagnostic Observation Schedule (ADOS), which was administered by trained clinicians to all study participants [Gotham, Pickles, & Lord, 2009]. We used the overall ADOS calibrated severity score (CSS), the social affect (SA) CSS, and the RRB CSS to highlight different domains of the ASD phenotype [Hus, Gotham, & Lord, 2014]. The ADOS CSS scores range from 1 to 10 with 10 indicating greatest severity and scores over 3 suggestive of an ASD diagnosis.

Cognitive ability was also considered in the analysis and was represented by verbal IQ and nonverbal IQ scores collected with the Wechsler Abbreviated Scale of Intelligence [Wechsler, 1999], the Differential Ability Scales [Elliot, 2007], or the Mullen Scales of Early Learning [Mullen, 1995], depending on the age and ability of participants. Similarly, the adaptive ability was collected and measured using the parent interview version of the Vineland Adaptive Behavior Scales (VABS), Second Edition [Sparrow, Balla, & Cicchetti, 2005] with a focus on the overall composite score in the current analysis.

Table 1. Prenatal Risk Factors and Perinatal Events

	16p11.2 deletion			16p11.2 duplication		
	Count	%	N	Count	%	N
<i>Prenatal risk factors</i>						
1. Advanced maternal age (>35 years)	29	42.0	69	18	27.3	66
2. Advanced paternal age (>40 years)	54	78.3	69	52	78.8	66
3. Preeclampsia	6	7.6	79	5	15.2	33
4. Use of antidepressants	5	7.0	71	2	6.5	31
5. Use of psychotropic medications	1	1.4	71	0	0.0	46
<i>Perinatal events</i>						
1. Low APGAR score at 5 min	2	4.5	44	2	12.5	16
2. Respiratory distress	20	24.1	83	6	17.6	34
3. Supplemental oxygen used	22	27.2	81	5	15.2	33
4. Ventilator used	5	6.3	80	1	2.9	35
5. Abnormal presentation	7	8.6	81	3	5.0	60
6. Preterm labor (<37 weeks of gestation)	18	22.8	79	6	17.6	34
7. C-section performed	29	31.5	92	18	25.0	72
8. Low birthweight (<2,500 g)	11	13.4	82	7	12.1	58

Cognitive and adaptive scores are reported with a mean of 100 and a *SD* of 15.

Given ADOS scores are not normalized as the cognitive and adaptive ability, scores from all measurements were normalized to a z-score for comparison across measurements.

Analytic Plan

Chi-square and independent two sample *t*-tests (of unequal variance) were conducted to compare initial demographics, independent variables, and outcome measures between 16p11.2 deletion and duplication carriers. To confirm independence between potential risk variables (sex, perinatal events, prenatal events, SNV w/CADD>20, and Rare CNV), two-tailed correlations were conducted and revealed no significant associations between variables across all participants (r 's < 0.24, P 's > 0.084) or within each carrier group (deletion carriers, r 's < 0.30, P 's > 0.10; duplication carriers, r 's < 0.34, P 's > 0.072) with one exception: Duplication carriers exhibited a positive relationship between the number of deleterious SNVs and number of rare CNV, $r = 0.59$, $P = 0.020$.

First, linear mixed-effects models were performed for each carrier group separately to assess the effect of all independent variables on measurements of ASD symptomatology (ADOS CSS, ADOS-SA CSS, and ADOS-RRB CSS) and cognitive and adaptive function (verbal IQ, nonverbal IQ, and VABS). Linear mixed-effects modeling was conducted in order to account for shared variance within the family by including a random intercept for each family. Sex was included in the analyses to ascertain contributions to phenotypic expression. Carrier group differences were directly tested within the second set of linear mixed effect models with identical parameters but an interaction with a group for each factor. All analyses were performed using SAS (version 9.4) using PROC MIXED with maximum likelihood and Satterthwaite denominator degrees of freedom. Standardized coefficients and 95% confidence intervals are provided in Figure 1. Additional post hoc analyses that involved repeating the analyses while controlling for critical factors (NVIQ for ASD symptomology model, ADOS CSS for cognitive and adaptive model) and for ascertainment status (initially identified proband vs. family member) are provided in Supplementary Materials and Figures S1 and S2.

Results

Table 2 describes the study sample; the final study population consisted of 173 individuals from 128 different families. The carrier groups differed in mean age (deletion

carriers significantly younger), the frequency of additional genetic events (higher for deletion carriers, though the equal number in probands only; see Table S1), the mean number of perinatal events (higher for deletion carriers), and the mean verbal IQ score (higher for duplication carriers). Deletion and duplication carriers had similar numbers of prenatal risk factors, similar proportions of male participants, and similar nonverbal IQ, Vineland composite, and ASD-symptom severity scores.

ASD Symptomatology

For deletion carriers, an increased number of perinatal events was associated with increased overall ASD symptoms (ADOS CSS: $t(61.4) = 2.08$, $P = 0.042$). Being female was associated with decreased restrictive and repetitive behaviors (ADOS-RRB CSS: $t(80) = -2.79$, $P = 0.007$). Three trends were noted: Related to secondary genetic hits, first, an increased number of SNVs showed a trend toward increased overall ASD severity (ADOS CSS: $t(81.2) = 1.97$, $P = 0.053$); and second, an increased number of rare CNVs showed a trend in association with increased restrictive and repetitive Behaviors (ADOS-RRB CSS: $t(80) = 1.93$, $P = 0.057$). Third, an increased number of perinatal events showed a trend towards increased social-affect deficits (ADOS-RRB CSS: $t(70.7) = 1.87$, $P = 0.065$).

For duplication carriers, female sex was associated with lower overall ASD symptomatology (ADOS CSS: $t(60) = -2.22$, $P = 0.031$) and decreased social impairment (ADOS-SA CSS: $t(58) = -2.92$, $P = 0.005$). One trend was noted: an increased number of rare CNVs was associated with increased ASD symptomatology (ADOS CSS: $t(60) = 1.82$, $P = 0.074$). No other variables were associated with changes in ASD symptomatology scores for duplication carriers.

Between carrier group differences related to the effects of the variables were noted, such that the effect of sex on social-affect deficits (reduced effect) was stronger for duplication females than deletion, $F(2, 138) = 4.39$, $P = 0.014$. In contrast, the effect of sex on RRBs (reduced effect) was stronger for deletion than duplication females, $F(2, 138) = 3.86$, $P = 0.023$. Lastly, the effect of perinatal events on social-affect deficits (increased effect) was stronger in deletion than duplication carriers, $F(2, 138) = 3.24$, $P = 0.043$.

Cognitive and Adaptive Function

For deletion carriers, increased number of perinatal events was associated with decreases in cognition and adaptive abilities (VIQ: $t(86) = -3.68$, $P < 0.001$; NVIQ: $t(86) = -3.4$, $P = 0.001$; VABS: $t(84) = -2.35$, $P = 0.021$). Additionally, increased number of rare CNVs was also associated with decreases in cognition (VIQ: $t(39.5) = -3.24$, $P = 0.002$; NVIQ: $t(43.9) = -2.92$, $P = 0.006$), but

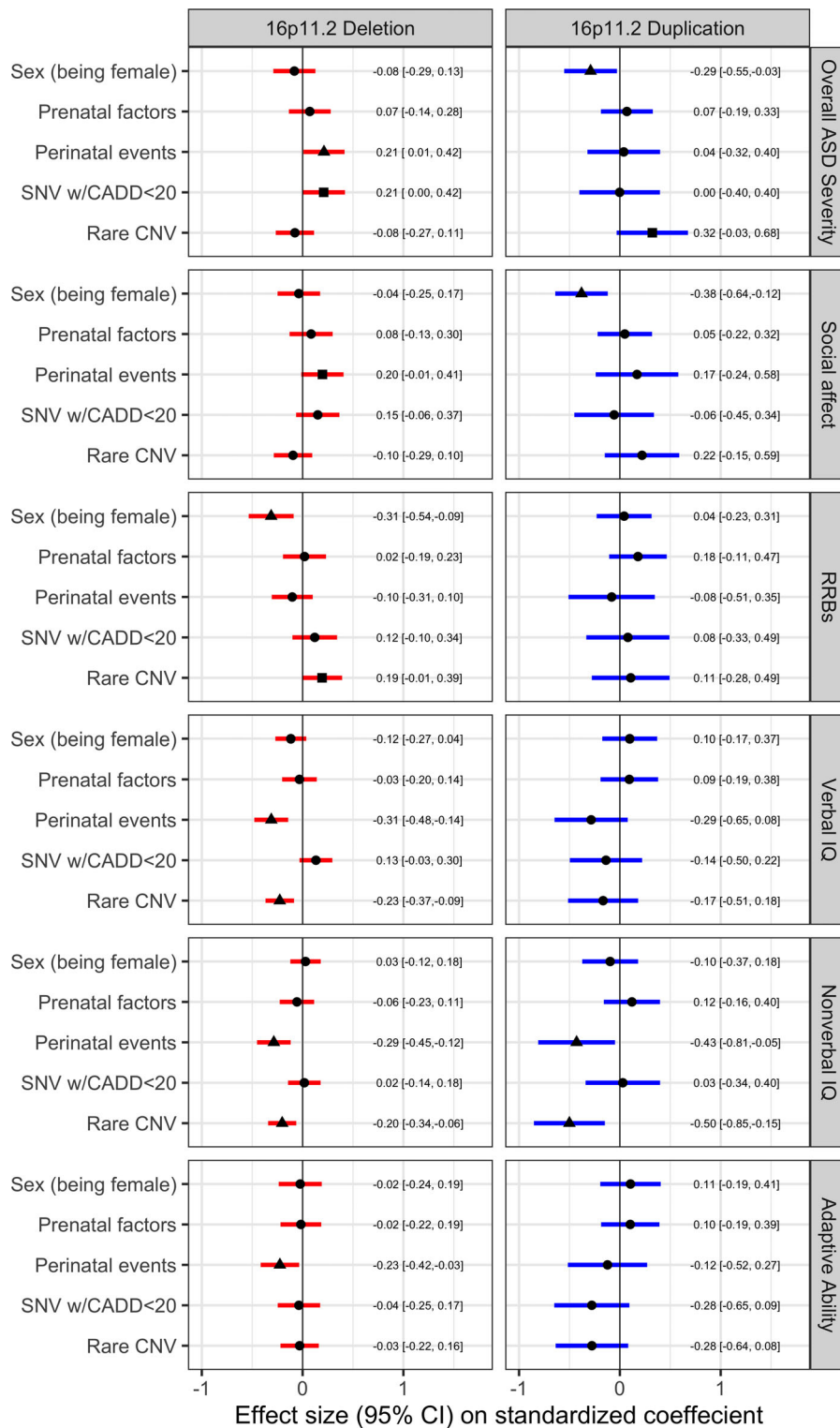


Figure 1. Effects of predictors on ASD symptomatology, cognition function, and adaptive function. Results of linear mixed-effects of the five independent variables (sex, perinatal events, prenatal events, SNV w/CADD>20, and rare CNV) for each of the outcome measurements of ASD symptomatology (ADOS CSS score, ADOS-SA CSS score, and ADOS-RRB CSS score), cognitive function (verbal IQ, nonverbal IQ), and adaptive function (Vineland Adaptive Behavior Composite Standard Score). Sex is designated as the effect of being female. The effect of a predictor variable is marked by a central marker, with its 95% confidence interval presented along a horizontal line. Triangular markers indicate a significant effect ($P < 0.05$), squares indicate a trend ($P < 0.1$), and circles indicate a nonsignificant effect.

Table 2. Descriptive Statistics for 16p11.2 Deletion and Duplication Carriers

Characteristics	Deletion	Duplication	P-value ^a
<i>Demographic</i>			
N this study	96	77	
Unique families N	86 (90%)	42 (54.5%)	
Age in years <i>M(SD)</i>	10.46 (9.32)	24.99 (19.30)	<0.001
Age range in years	0.8–48	1.7–80.1	
Male N (%)	73 (53%)	40 (52%)	0.88
Clinical ASD diagnosis N(%)	22 (22.9%)	10 (14.5%)	0.25
<i>Predictor variables (continuous) - Mean number of events (SD)</i>			
Rare CNV	1.1 (1.3)	0.56 (1.0)	0.003
SNV w/CADD > 20	0.84 (1.0)	0.49 (0.84)	0.014
Prenatal risk factors	1.1 (0.87)	1.1 (0.81)	0.88
Perinatal events	1.2 (1.7)	0.67 (1.2)	0.018
<i>Genetic factor predictor variables as categorical N = yes(%)</i>			
Both rare CNV and SNV	38 (40%)	17 (22%)	<0.001
Rare CNV only	18 (19%)	6 (8%)	
SNV w/CADD > 20	12 (13%)	6 (8%)	
No rare CNV or SNV	28 (29%)	48 (62%)	
<i>Pre- and Peri-natal predictor variables as categorical N = yes(%)</i>			
Both pre- and peri-natal factors	30 (31%)	18 (23%)	0.20
Prenatal risk factors only	18 (19%)	9 (12%)	
Perinatal events only	31 (32%)	36 (47%)	
No pre- or peri-natal factors	17 (18%)	14 (18%)	
<i>ASD Symptoms via ADOS CSS^b - M(SD)</i>			
Overall	3.2 (2.3)	3.0 (2.4)	0.56
Social affect (SA)	3.5 (2.3)	3.3 (2.5)	0.55
Restrictive and repetitive behaviors (RRBs)	3.7 (2.8)	3.5 (2.6)	0.77
<i>Cognitive and adaptive functioning - M(SD)^c</i>			
Verbal IQ	79.4 (18.4)	87.0 (23.9)	0.023
Nonverbal IQ	87.5 (16.7)	84.8 (23.1)	0.38
Vineland Adaptive Behavior Scales (VABS)	78.7 (13.1)	81.0 (15.6)	0.31

^aOne-way analysis of variance (of unequal variance) conducted for comparison of means and χ^2 test conducted for comparison of percentages. All significant P-values < .05 are given in italics.

^bADOS Calibrated Severity Score, standardized 1–10 scoring with 1–3 indicating non-ASD and 4–10 indicating ASD.

^cVerbal IQ, nonverbal IQ, and Vineland scores are standardized to a mean = 100, SD = 15.

not adaptive abilities (VABS: $t(84) = -0.31$, $P = 0.79 = 6$). No other variables were associated with changes in cognitive and adaptive function for deletion carriers.

For duplication carriers, none of the independent variables was associated with changes in standardized verbal IQ scores or adaptive skills. Similar to deletion carriers, decreased nonverbal cognition was associated with increased number of rare CNVs (NVIQ: $t(62.9) = -2.25$, $P = 0.028$) and an increased number of perinatal events (NVIQ: $t(62.5) = -2.84$, $P = 0.006$).

Between carrier group differences related to the effects of the variables were noted (see Fig. S2), such that the estimated effects of increased perinatal events on verbal and nonverbal cognition (reduced effect) was stronger for duplication than deletion carriers [VIQ: $F(2,130) = 5.07$, $P = 0.0076$; NIVQ: $F(2,136) = 6.01$, $P = 0.0032$]. The estimated effect of increased rare CNVs was stronger for deletion carriers in verbal cognition (VIQ: $F(2,142) = 4.81$, $P = 0.0095$) but stronger for duplication carriers in nonverbal cognition (NVIQ: $F(2,145) = 7.24$, $P = 0.001$).

Discussion

This study investigated the impact of secondary genetic events, prenatal and perinatal environmental factors, and sex on phenotypic heterogeneity among 16p11.2 CNV carriers. Critically, despite a lack of group-level differences in ASD symptomatology, nonverbal cognition, and adaptive functioning between carrier groups, we found differential contributions of our risk factors for each carrier group. Broadly, we identified patterns suggesting that overall ASD symptomatology was influenced by the frequency of perinatal events in deletion carriers. ASD symptomatology was influenced by sex in duplication carriers, such that those of female sex showed significantly less ASD-related deficits, specifically within social-affect symptoms, whereas female deletion carriers exhibited restricted and repetitive behaviors. The frequency of perinatal events and the presence of additional CNVs conferred additional deficits in nonverbal cognition in both deletion and duplication carriers, with increased perinatal

events being associated with reduced verbal cognition and adaptive abilities in deletion carriers.

Perinatal contributions. Our results support previous reports of the interactive effects of perinatal events and CNVs on increased ASD symptomatology and reduced cognitive and adaptive behaviors [Mazina et al., 2014], specifically for 16p11.2 deletion carriers. Importantly, the effect of perinatal events remained in models controlling for nonverbal cognition (see Fig. S1). Deletion carriers had significantly more perinatal events (40% deletion vs. 22% duplication), which lends greater variability and power to detect relationships between risk factors and outcomes. The lack of a perinatal contribution to ASD symptomatology in duplication carriers may be related to the reduced severity phenotype commonly observed [Steinman et al., 2016; Niarchou et al., 2019a; Niarchou et al., 2019b], yet, we caution over-interpretation of the lack of an association, given the limited sample size of this exploratory study. In addition, although an item-level analysis was beyond the scope of this study, continued work should specifically investigate high-frequency perinatal events: respiratory distress, supplemental oxygen used, preterm labor, and Caesarean section performed. These elements have been previously noted as risk factors for both ASD and intellectual disability [see Modabbernia et al., 2016 for a review], and it is likely that these events stem from a common underlying cause. However, the relative contribution of each event, as well as patterns related to certain constellations of perinatal events, should be explored in future studies.

Secondary genetic contributions. While rare CNVs were contributing factors of cognition for both deletion carriers (verbal, nonverbal) and duplication carriers (nonverbal only), secondary genetic factors were weaker in regard to ASD symptomatology. Two trends within the deletion-carrier group indicated that deleterious SNV and rare CNVs may impact overall ASD severity and RRBs, respectively. In addition, when cognitive models controlled for ASD severity (via ADOS CSS, see Fig. S2), an increased number of additional rare CNVs accounted for reduced verbal and nonverbal cognition. These findings are aligned with prior work emphasizing a strong role of more severe mutations (e.g., de novo mutations, rare CNVs, and deleterious SNVs) on intellectual and cognitive capacity [Girirajan et al., 2010; Hamdan et al., 2014].

There is clearly a complex phenotype–16p11.2 genotype relationship with recent work suggesting that 16p11.2 deletions may lead to intellectual impairments directly, yet ASD indirectly (i.e., ASD due to reduced cognitive abilities) [Moreno-De-Luca, Moreno-De-Luca, Cubells, & Sanders, 2014]. However, although females with ASD show a higher mutational burden and lower cognitive abilities, adjustments for cognition do not

change the excess mutational burden [Jacquemont et al., 2014], indicating that these effects are not solely related to cognition. Moving forward toward clinical trials, it will be important to emphasize cross-level interactions to understand the mechanism by which cognitive abilities may be impacted by secondary genetic factors. For instance, evidence from neuroimaging suggests divergent mechanisms by which 16p11.2 carriers process information [Hudac et al., 2015], which may further describe how cognitive abilities may support (or interfere with) the social phenotype, broadly, and more specifically, ASD symptomatology.

As a caveat, prior work indicated a similar number of secondary de novo CNVs, often regarded as more pathogenic, in 16p11.2 probands specifically [Duyzend et al., 2016]. Although our sample seemingly illustrates larger numbers of potentially disruptive genetic events than duplication carriers (71% deletion vs. 38% duplication, see Table 2), these effects may be inflated due to the lack of additional events in family members. When we specifically examine rates of potentially disruptive genetic events in the initially identified probands (Table S1), there are a similar number of rates between deletion and duplication probands (81% deletion and 88% duplication). However, results in probands only (Fig. S2) support the hypothesis that additional genetic factors are contributing to cognitive and adaptive functioning, in particular.

Female protective effect. We also found associations between female sex and less severe ASD symptomatology among both 16p11.2 deletion carriers (RRB subdomain) and duplication carriers (overall and SA subdomain). This finding adds additional support for a female protective effect [Robinson, Lichtenstein, Anckarsäter, Happé, & Ronald, 2013] that describes the phenomenon in which females with ASD exhibit less severe ASD symptomatology and is less likely to be diagnosed than males. Importantly, sex did not contribute to cognitive or adaptive abilities, suggesting that the female protective effect is specific to ASD symptomatology. Previous work using clinical samples has shown that females with diagnoses of ASD have lower IQs and are more severely affected than their male counterparts [Fombonne, 2003]. Within our sample, IQ did not vary based upon sex within carrier group (post hoc, P 's > 0.10) or as related to ASD diagnosis (post hoc, P 's > 0.17), though a trend indicated reduced nonverbal cognition in female duplication carriers with ASD (post hoc, P 's = 0.061). This is related to the analysis indicating the female protective effect for nonverbal cognition in duplications when overall ASD symptomatology was controlled (Fig. S1).

Our primary data indicate a reduction of ASD symptoms for females in both carrier groups, yet, there may be additional implications for being female and having

reduced nonverbal cognition. For one, although our study did not consider the interaction between environmental and genetic factors, it is important to begin to understand how the constellation of cross-factors relates to the 16p11.2 phenotype. Robinson et al. [2014] found that females in their clinically based sample had nearly twice the number of loss of function mutations than males, suggesting that females may require a greater etiological burden to manifest greater ASD symptomatology [Robinson et al., 2013].

Future directions. The five risk factors accounted for 9 to 20% of the variability (R^2 values) in the phenotypic outcome measurements. These findings suggest that while the additional factors that we identified are contributing to the variability in ASD symptoms there are other factors that may account for this variance.

Previous work indicates the strong role of parental background in accounting for this variance, including contributions from noncarrier parents [Moreno-De-Luca et al., 2015], suggesting a potential dimensional shift related to underlying genetic background. A random intercept was included in this analysis as a way to control for multiple family members, consistent with a previous study that did not find evidence of correspondence of cognitive abilities or social skills between carrier children and carrier parents, [Snyder et al., 2016]. However, we did not explicitly examine relationships between first-degree family members (e.g., initially identified child proband compared to carrier parent, noncarrier parent, and/or biparental mean). It is likely that additional familial factors at both the genetic or environmental levels are contributing to the phenotype. For instance, noncarrier family members exhibit anatomical brain alterations compared to noncarrier nonfamily controls [Martin-Brevet et al., 2018], suggesting a more complex investigation of the possible parental background and additional risk factors is critical to better account for variability.

There are several limitations to note. First, in this study, we examined whether the quantity of prenatal and perinatal events was related to functional outcomes without consideration of specific events. It may be the case that the severity of certain events outweighs other events, which may be evidenced by contrasting findings in the extant literature. For one, an earlier meta-analysis found no association between preeclampsia and ASD risk [Gardener et al., 2009], in contrast with a more recent meta-analysis [Dachew et al., 2018] that indicated preeclampsia as a strong ASD-risk factor. Second, we are limited in our ability to fully assess codependent or shared aspects, such that the factors targeted by these analyses may reflect underlying biology and/or environmental features. For instance, it is difficult to determine prenatal risk factors and perinatal events are truly independent, or rather, may reflect a common unified consequence of

interrelated factors (e.g., women with advanced maternal age are more likely to give birth via C-section). Relatedly, several key demographic features were not addressed within our analyses. Of particular note, due to cascade genetic testing commonly used to confirm and subsequently identify additional familial 16p11.2 carriers, a larger proportion of the duplication carriers were related, older adults (see Table 2). These features may have led to increased variance between carriers (i.e., larger confidence intervals), though post hoc analyses with only initially identified probands revealed similar findings (see Fig. S2). Third, our study is exploratory in nature, such that continued work is required to confirm that these differences are not mere detection differences, but instead reflective of underlying mechanisms stemming from 16p11.2 CNVs. Here, effect sizes are provided as confidence intervals around standardized coefficients in order to better represent relative contributions across different factors. However, these values are not meant to be clinically significant but rather descriptive, as more extensive work with larger samples would be required to determine statistical reliability and thus, clinical relevance.

Overall, this study demonstrated support for the contribution of ASD-associated risk factors (perinatal events, sex, and secondary rare CNVs) to increased ASD symptoms and cognitive/adaptive impairment in individuals with 16p11.2 CNVs. These findings are consistent with prior work suggesting that it is the confluence of multiple genetic risk factors that increases the likelihood of neurodevelopmental disorders, more broadly [Girirajan et al., 2012]. Interactions with environmental factors related to social and psychological familial aspects should also be considered [Baker, Devine, Ng-Cordell, Raymond, & Hughes, 2020] as a means to evaluate and understand the gene by environment mechanisms [Wender & Veenstra-Vanderweele, 2017]. As a note of caution, this exploratory study serves as a first foray into specifying specific secondary genetic and environmental factors related to the 16p11.2 CNV phenotype. Future work would benefit from a more careful exploration of the presence of specific environmental and genetic factors.

Acknowledgments

This work was supported by a grant from the Simons Foundation (SFARI award #198677 to RAB, EH, RPG-K, and WKC). We are grateful to all of the families at the participating Simons Variation in Individuals Project (Simons VIP) sites, as well as the Simons VIP working group (Simons VIP consortium, *Neuron*, 73(6): 1063-1067, 2012). We appreciate obtaining access to phenotypic data on SFARI Base. Approved researchers can obtain the Simons VIP population dataset described in this study by applying at <https://base.sfari.org>. The Simons VIP Consortium includes: H. Alupay,

B. Aaronson, S. Ackerman, K. Ankenmann, C. Atwell, E. Aylward, A. Beaudet, M. Benedetti, J. Berman, R. Bernier, A. Bibb, L. Blaskey, C. Brewton, R. Buckner, P. Bukshpun, J. Burko, B. Cerban, Q. Chen, M. Cheong, Z. Chu, W. Chung, C. Dale, A. Dempsey, J. Elgin, J. Olson, Y. Evans, W. A. Faucett, G. Fischbach, S. Garza, J. Gerds, S. Gobuty, R. Goin-Kochel, P. E. Grant, L. Green Snyder, M. Greenup, E. Hanson, K. Hines, L. Hinkley, J. Hunter, R. Jeremy, K. Johnson, S. Kanne, S. Kessler, S. Khan, A. Laakman, M. Lasala, D. Ledbetter, H. Lee, C. Lese Martin, A. Lian Cavanagh, A. Llorens, T. Luks, E. Marco, A. Martin, G. Marzano, K. McGovern, R. McNally Keehn, D. Miller, F. Miller, T. Moss, P. Mukherjee, S. Nagarajan, K. Nowell, J. Owen, A. Paal, A. Packer, P. Page, B. Paul, N. Pojman, M. Proud, S. Qasmieh, M. Ramocki, B. Reilly, T. Roberts, D. Shaw, E. Sherr, T. Sinha, B. Smith-Packard, A. Snow, S. Spence, J. Spiro, K. Steinman, A. Stevens, V. Swarnakar, J. Tjernagel, C. Triantafallou, R. Vaughan, N. Visyak, M. Wakahiro, T. Ward, and J. Wenegrat.

Conflict of interest

E.E.E. is an investigator of the Howard Hughes Medical Institute. E.E.E. is on the scientific advisory board (SAB) of DNAnexus, Inc. There are no other conflicts of interest to report.

References

Baker, K., Devine, R. T., Ng-Cordell, E., Raymond, F. L., & Hughes, C. (2020). Childhood intellectual disability and parents' mental health: integrating social, psychological and genetic influences. *The British Journal of Psychiatry*. <https://doi.org/10.1192/bjp.2020.38>.

Chen, S. W., Zhong, X. S., Jiang, L. N., Zheng, X. Y., Xiong, Y. Q., Ma, S. J., ... Chen, Q. (2016). Maternal autoimmune diseases and the risk of autism spectrum disorders in offspring: A systematic review and meta-analysis. *Behavioural Brain Research*, 296, 61–69.

Chien, Y. L., Chou, M. C., Chou, W. J., Wu, Y. Y., Tsai, W. C., Chiu, Y. N., & Gau, S. S. F. (2019). Prenatal and perinatal risk factors and the clinical implications on autism spectrum disorder. *Autism*, 23(3), 783–791.

Coe, B. P., Stessman, H. A., Sulovari, A., Geisheker, M. R., Bakken, T. E., Lake, A. M., ... Eichler, E. E. (2019). Neurodevelopmental disease genes implicated by de novo mutation and copy number variation morbidity. *Nature Genetics*, 51(1), 106–116.

Constantino, J. N., & Charman, T. (2012). Gender bias, female resilience, and the sex ratio in autism. *Journal of the American Academy of Child & Adolescent Psychiatry*, 51(8), 756–758.

Dachew, B. A., Mamun, A., Maravilla, J. C., & Alati, R. (2018). Pre-eclampsia and the risk of autism-spectrum disorder in

offspring: meta-analysis. *The British Journal of Psychiatry*, 212, 142–147.

Danecek, P., Auton, A., Abecasis, G., Albers, C. A., Banks, E., DePristo, M. A., ... 1000 Genomes Project Analysis Group. (2011). The variant call format and VCFtools. *Bioinformatics*, 27, 2156–2158.

D'Angelo, D., Lebon, S., Chen, Q., Martin-Brevet, S., Snyder, L. G., Hippolyte, L., ... Pain, A. (2016). Defining the effect of the 16p11.2 duplication on cognition, behavior, and medical comorbidities. *JAMA Psychiatry*, 73(1), 20–30.

Duyzend, M. H., Nuttle, X., Coe, B. P., Baker, C., Nickerson, D. A., Bernier, R., & Eichler, E. E. (2016). Maternal modifiers and parent-of-origin bias of the autism-associated 16p11.2 cnv. *The American Journal of Human Genetics*, 98(1), 45–57.

Elliot, C. D. (2007). *Differential ability scales* (2nd ed.). San Antonio, TX: Harcourt Assessment.

Fombonne, E. (2003). Epidemiological surveys of autism and other pervasive developmental disorders: An update. *Journal of Autism and Developmental Disorders*, 33(4), 365–382.

Gardener, H., Spiegelman, D., & Buka, S. L. (2009). Prenatal risk factors for autism: Comprehensive meta-analysis. *The British Journal of Psychiatry*, 195(1), 7–14.

Gardener, H., Spiegelman, D., & Buka, S. L. (2011). Perinatal and neonatal risk factors for autism: A comprehensive meta-analysis. *Pediatrics*, 128(2), 344–355.

Garrison, E. (2012). Vcflib: A C++ library for parsing and manipulating VCF files. GitHub <https://github.com/ekg/vcflib>.

Girirajan, S., Rosenfeld, J. A., Coe, B. P., Parikh, S., Friedman, N., Goldstein, A., ... Nezarati, M. M. (2012). Phenotypic heterogeneity of genomic disorders and rare copy-number variants. *New England Journal of Medicine*, 367(14), 1321–1331.

Girirajan, S., Rosenfeld, J. A., Cooper, G. M., Antonacci, F., Siswara, P., Itsara, A., ... Mefford, H. C. (2010). A recurrent 16p12.1 microdeletion supports a two-hit model for severe developmental delay. *Nature Genetics*, 42(3), 203.

Gotham, K., Pickles, A., & Lord, C. (2009). Standardizing ADOS scores for a measure of severity in autism spectrum disorders. *Journal of Autism and Developmental Disorders*, 39(5), 693–705.

Guo, H., Wang, T., Wu, H., Long, M., Coe, B. P., Li, H., ... Bai, T. (2018). Inherited and multiple de novo mutations in autism/developmental delay risk genes suggest a multifactorial model. *Molecular Autism*, 9(1), 64.

Hamdan, F. F., Srouf, M., Capo-Chichi, J. M., Daoud, H., Nassif, C., Patry, L., ... Henrion, E. (2014). De novo mutations in moderate or severe intellectual disability. *PLoS Genetics*, 10(10), e1004772.

Hanson, E., Bernier, R., Porche, K., Jackson, F. I., Goin-Kochel, R. P., Snyder, L. G., ... Chen, Q. (2015). The cognitive and behavioral phenotype of the 16p11.2 deletion in a clinically ascertained population. *Biological Psychiatry*, 77(9), 785–793.

Hisle-Gorman, E., Susi, A., Stokes, T., Gorman, G., Erdie-Lalena, C., & Nylund, C. M. (2018). Prenatal, perinatal, and neonatal risk factors of autism spectrum disorder. *Pediatric Research*, 84(2), 190–198.

Hudac, C. M., Kresse, A., Aaronson, B., DesChamps, T. D., Webb, S. J., & Bernier, R. A. (2015). Modulation of mu attenuation to social stimuli in children and adults with 16p11.2 deletions and duplications. *Journal of Neurodevelopmental Disorders*, 7(1), 25.

- Hus, V., Gotham, K., & Lord, C. (2014). Standardizing ADOS domain scores: Separating severity of social affect and restricted and repetitive behaviors. *Journal of Autism and Developmental Disorders*, 44(10), 2400–2412.
- Iossifov, I., O’Roak, B. J., Sanders, S. J., Ronemus, M., Krumm, N., Levy, D., ... Smith, J. D. (2014). The contribution of de novo coding mutations to autism spectrum disorder. *Nature*, 515(7526), 216–221.
- Jacquemont, S., Coe, B. P., Hersch, M., Duyzend, M. H., Krumm, N., Bergmann, S., ... Eichler, E. E. (2014). A higher mutational burden in females supports a “female protective model” in neurodevelopmental disorders. *The American Journal of Human Genetics*, 94(3), 415–425.
- Jiang, H. Y., Xu, L. L., Shao, L., Xia, R. M., Yu, Z. H., Ling, Z. X., ... Ruan, B. (2016). Maternal infection during pregnancy and risk of autism spectrum disorders: A systematic review and meta-analysis. *Brain, Behavior, and Immunity*, 58, 165–172.
- Kaplan, Y. C., Keskin-Arslan, E., Acar, S., & Sozmen, K. (2016). Prenatal selective serotonin reuptake inhibitor use and risk of autism spectrum disorder in the children: A systematic review and meta-analysis. *Reproductive Toxicology*, 60, 174.
- Kircher, M., Witten, D. M., Jain, P., O’Roak, B. J., Cooper, G. M., & Shendure, J. (2014). A general framework for estimating the relative pathogenicity of human genetic variants. *Nature Genetics*, 46, 310–315.
- Kirov, G., Rees, E., Walters, J. T. R., Escott-Price, V., Georgieva, L., Richards, A. L., ... Owen, M. J. (2014). The penetrance of copy number variations for schizophrenia and developmental delay. *Biological Psychiatry*, 75(5), 378–385.
- Kreiser, N. L., & White, S. W. (2014). ASD in females: Are we overstating the gender difference in diagnosis? *Clinical Child and Family Psychology Review*, 17(1), 67–84.
- Krumm, N., Turner, T. N., Baker, C., Vives, L., Mohajeri, K., Witherspoon, K., ... Leal, S. M. (2015). Excess of rare, inherited truncating mutations in autism. *Nature Genetics*, 47(6), 582–588.
- Kumar, R. A., KaraMohamed, S., Sudi, J., Conrad, D. F., Brune, C., Badner, J. A., ... Christian, S. L. (2008). Recurrent 16p11.2 microdeletions in autism. *Human Molecular Genetics*, 17(4), 628–638.
- Lai, M. C., Lombardo, M. V., Auyeung, B., Chakrabarti, B., & Baron-Cohen, S. (2015). Sex/gender differences and autism: setting the scene for future research. *Journal of the American Academy of Child & Adolescent Psychiatry*, 54(1), 11–24.
- Langridge, A. T., Glasson, E. J., Nassar, N., Jacoby, P., Pennell, C., Hagan, R., ... Stanley, F. J. (2013). Maternal conditions and perinatal characteristics associated with autism spectrum disorder and intellectual disability. *PLoS One*, 8(1), e50963.
- Martin-Brevet, S., Rodríguez-Herreros, B., Nielsen, J. A., Moreau, C., Modenato, C., Maillard, A. M., ... Zürcher, N. R. (2018). Quantifying the effects of 16p11.2 Copy number variants on brain structure: A multisite genetic-first study. *Biological Psychiatry*, 84(4), 253–264.
- Matelski, L., & Van de Water, J. (2016). Risk factors in autism: Thinking outside the brain. *Journal of Autoimmunity*, 67, 1–7.
- Mazina, V., Gerdt, J., Trinh, S., Ankenman, K., Ward, T., Dennis, M. Y., ... Bernier, R. (2014). Epigenetics of autism-related impairment: Copy number variation and maternal infection. *Journal of Developmental and Behavioral Pediatrics*, 36(2), 61–67.
- McCarthy, S. E., Makarov, V., Kirov, G., Addington, A. M., McClellan, J., Yoon, S., ... Krause, V. (2009). Microduplications of 16p11.2 are associated with schizophrenia. *Nature Genetics*, 41(11), 1223–1227.
- Modabbernia, A., Mollon, J., Boffetta, P., & Reichenberg, A. (2016). Impaired gas exchange at birth and risk of intellectual disability and autism: A meta-analysis. *Journal of Autism and Developmental Disorders*, 46(5), 1847–1859.
- Moreno-De-Luca, A., Evans, D. W., Boomer, K. B., Hanson, E., Bernier, R., Goin-Kochel, R. P., ... Hare, A. E. (2015). The role of parental cognitive, behavioral, and motor profiles in clinical variability in individuals with chromosome 16p11.2 deletions. *JAMA Psychiatry*, 72(2), 119–126.
- Moreno-De-Luca, D., Moreno-De-Luca, A., Cubells, J. F., & Sanders, S. J. (2014). Cross-disorder comparison of four neuropsychiatric CNV loci. *Current Genetic Medicine Reports*, 2(3), 151–161.
- Mullen, E. M. (1995). *Mullen scales of early learn: AGS edition*. Minneapolis, MN: Pearson (AGS).
- Niarchou, M., Chawner, S. J., Doherty, J. L., Maillard, A. M., Jacquemont, S., Chung, W. K., ... Linden, D. E. (2019a). Psychiatric disorders in children with 16p11.2 deletion and duplication. *Translational Psychiatry*, 9(1), 1–8.
- O’Roak, B. J., Deriziotis, P., Lee, C., Vives, L., Schwartz, J. J., Girirajan, S., ... Rieder, M. J. (2011). Exome sequencing in sporadic autism spectrum disorders identifies severe de novo mutations. *Nature Genetics*, 43(6), 585–589.
- Park, H. R., Lee, J. M., Moon, H. E., Lee, D. S., Kim, B. N., Kim, J., ... Paek, S. H. (2016). A short review on the current understanding of autism spectrum disorders. *Experimental Neurobiology*, 25(1), 1–13.
- Perrone-McGovern, K., Simon-Dack, S., & Niccolai, L. (2015). Prenatal and perinatal factors related to autism, IQ, and adaptive functioning. *The Journal of Genetic Psychology*, 176(1), 1–10.
- Petrovski, S., Wang, Q., Heinzen, E. L., Allen, A. S., & Goldstein, D. B. (2013). Genic intolerance to functional variation and the interpretation of personal genomes. *PLoS Genetics*, 9, e1003709.
- Polyak, A., Rosenfeld, J. A., & Girirajan, S. (2015). An assessment of sex bias in neurodevelopmental disorders. *Genome Medicine*, 7(94), 1–11.
- Rais, T. B., & Rais, A. (2014). Association between antidepressants use during pregnancy and autistic spectrum disorders: A meta-analysis. *Innovations in Clinical Neuroscience*, 11(5–6), 18–22.
- Robinson, E. B., Lichtenstein, P., Anckarsäter, H., Happé, F., & Ronald, A. (2013). Examining and interpreting the female protective effect against autistic behavior. *Proceedings of the National Academy of Sciences*, 110(13), 5258–5262.
- Robinson, E. B., Samocha, K. E., Kosmicki, J. A., McGrath, L., Neale, B. M., Perlis, R. H., & Daly, M. J. (2014). Autism spectrum disorder severity reflects the average contribution of de novo and familial influences. *Proceedings of the National Academy of Sciences*, 111(42), 15161–15165.
- Rosenfeld, J. A., Coe, B. P., Eichler, E. E., Cuckle, H. D., & Shaffer, L. G. (2013). Estimates of penetrance for recurrent

- pathogenic copy-number variations. *Genetics in Medicine*, 15(6), 478–481.
- Samocha, K. E., Robinson, E. B., Sanders, S. J., Stevens, C., Sabo, A., McGrath, L. M., ... Wall, D. P. (2014). A framework for the interpretation of de novo mutation in human disease. *Nature Genetics*, 46(9), 944–950.
- Sanders, S. J., He, X., Willsey, A. J., Ercan-Sencicek, A. G., Samocha, K. E., Cicek, A. E., ... Goldberg, A. P. (2015). Insights into autism spectrum disorder genomic architecture and biology from 71 risk loci. *Neuron*, 87(6), 1215–1233.
- Schrieken, M., Visser, J., Oosterling, I., van Steijn, D., Bons, D., Draaisma, J., ... Rommelse, N. (2013). Head circumference and height abnormalities in autism revisited: The role of pre- and perinatal risk factors. *European Child & Adolescent Psychiatry*, 22(1), 35–43.
- Simons VIP Consortium. (2012). Simons Variation in Individuals Project (Simons VIP): A genetics-first approach to studying autism spectrum and related neurodevelopmental disorders. *Neuron*, 73(6), 1063–1067.
- Snyder, L. G., D'Angelo, D., Chen, Q., Bernier, R., Goin-Kochel, R. P., Wallace, A. S., ... Kuschner, E. (2016). Autism spectrum disorder, developmental and psychiatric features in 16p11.2 duplication. *Journal of Autism and Developmental Disorders*, 46, 2734–2748.
- Sparrow, S. S., Balla, D. A., & Cicchetti, D. V. (2005). Vineland-II adaptive behavior scales. Circle Pines, MN: Pearson Assessments.
- Steinman, K. J., Spence, S. J., Ramocki, M. B., Proud, M. B., Kessler, S. K., Marco, E. J., ... Sherr, E. H. (2016). 16p11.2 deletion and duplication: characterizing neurologic phenotypes in a large clinically ascertained cohort. *American Journal of Medical Genetics, Part A*, 170(11), 2943–2955.
- Visser, J. C., Rommelse, N., Vink, L., Schrieken, M., Oosterling, I. J., van der Gaag, R. J., & Buitelaar, J. K. (2013). Narrowly versus broadly defined autism spectrum disorders: Differences in pre- and perinatal risk factors. *Journal of Autism and Developmental Disorders*, 43(7), 1505–1516.
- Wechsler, D. (1999). Wechsler abbreviated scale of intelligence. San Antonio, TX: The Psychological Corporation.
- Weiss, L. A., Shen, Y., Korn, J. M., Arking, D. E., Miller, D. T., Fossdal, R., ... Platt, O. S. (2008). Association between microdeletion and microduplication at 16p11.2 and autism. *New England Journal of Medicine*, 358(7), 667–675.
- Wender, C. L., & Veenstra-VanderWeele, J. (2017). Challenge and potential for research on gene-environment interactions in autism spectrum disorder. In *Gene-environment transactions in developmental psychopathology* (pp. 157–176). Cham: Springer.
- Zufferey, F., Sherr, E. H., Beckmann, N. D., Hanson, E., Maillard, A. M., Hippolyte, L., ... Aylward, E. (2012). A 600 kb deletion syndrome at 16p11.2 leads to energy imbalance and neuropsychiatric disorders. *Journal of Medical Genetics*, 49(10), 660–668.

Supporting Information

Additional supporting information may be found online in the Supporting Information section at the end of the article.

Appendix S1: Supporting Information

Table S1 Sequenced Individuals

Table S2: Rare CNV calls (frequency < 0.1% controls) in the 16p11.2 probands screened

Table S3: Private SNV calls